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Activity of HIV-I Protease Inhibitors in Various Enzymatic Assays.

J.Roesel,B.Poncioni,E.Alteri,J.Wood,G.Bold,A.Faessler,
H.Rueegger,M.Lang,P.Schneider

Pharmaceutical Research Department, CIBA-GEIGY Ltd., Basel,
Switzerland.

Retroviral proteases have been identified as useful targets for antiviral interference. Their main function is to allow correct processing of viral structural proteins; thereby ensuring the maturation of infectious progeny virus. Based on its amino acid sequence and structural conformation, HIV-I protease has been identified as an aspartic protease. This enabled us to synthesize HIV-I protease specific inhibitors. These inhibitors were characterized in cleavage assays using various proteases, peptides and viral precursor protein.

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Subcellular Localization and Antiviral Activity of Cationic Dye/Poly r(A-U) Combinations.
K Krabill, JM Jamison, J Gilloteaux, DG Flowers and C-c Tsai. Dept of Chemistry, Kent State Univ, Kent, OH 44242 and Dept of Anatomy, NE Ohio Univ COM, Rootstown, OH 44272 USA.

Experiments have been designed to examine the effects of 24 anthraquinone (AQ), xanthene (XAN) and ellipticine (EPC) dyes on the antiviral and interferon-inducing activities of poly r(A-U) using a human foreskin fibroblast - vesicular stomatitis virus bioassay in which 0.05 mM dye has been combined with poly r(A-U) to produce a dye/ribonucleotide ratio of 1/4. Poly r(A-U) and the dyes alone were not effective antiviral agents. When poly r(A-U) was combined with adriamycin (ADR), ametantrone (HAQ), carminic acid (CAR), daunomycin (DMN), mitoxantrone (DHAQ), N²-methyl-9-hydroxy-ellipticine (NMHE), N²,N⁶-dimethyl-9-hydroxy-ellipticine (DMHE), rhodamine 123 (R123), rhodamine B (RB), rhodamine 6G (R6G) and sulforhodamine B (SB), antiviral activity of poly r(A-U) was enhanced 8- to 20-fold, while 50% effective doses of the poly r(A-U), ADR, CAR, DMN, NMHE, HAQ, R6G, RB, DHAQ, DMHE, SB and R123 decreased 18-, 60-, 60-, 61-, 113-, 125-, 156-, 250-, 251-, 274-, 313- and 347-fold, respectively. Additional bioassays demonstrated that the enhanced antiviral activity of the dye/poly r(A-U) combinations was not due to increased interferon induction, direct viral inactivation or cytotoxicity. Bright field and fluorescence micrographs of human foreskin fibroblast monolayers co-incubated with active members of each of the 3 families of dyes, poly r(A-U) alone or with the dye/poly r(A-U) combinations exhibited three basic patterns: 1. the dye entered the cell as well as the nucleus and the accumulation of the dye in the nucleus was greatly potentiated by the poly r(A-U), (AQ); 2. the dye entered the cell and was sequestered in the cytoplasm, but readily accumulated in the nucleus in the presence of poly r(A-U), (EPC); 3. the dye entered the cell and was sequestered in cytoplasmic organelles (mitochondria) and only the poly r(A-U) and minute quantities of the dye accumulated in the nucleus when the dye was combined with poly r(A-U) (XAN). These results suggested that the potentiated antiviral activity of the AQ (or EPC)/poly r(A-U) combinations was due to the altered kinetics of uptake and subcellular distribution and the subsequent modulation of one or more nuclear (nucleolar) processes. Xanthene dyes have been shown to exert a cytostatic effect on cultured cells due to their disruption of mitochondrial energy production and the subsequent inability of cells to attain a critical content of essential components, such as rRNA, necessary for cell entrance into the pre-replicative compartment of G₁ phase. Thus it appears that xanthene dyes potentiated the antiviral activity of poly r(A-U) by interfering with cellular processes.